

**New CLSI/NCCLS Antimicrobial Susceptibility Testing (AST) Recommendations  
M100-S15 Checklist**

NA, not applicable

Will Implement	Previously Implemented	NA	Action
<b>General</b>			
			Review current <b>product insert</b> from commercial antimicrobial susceptibility testing products used in my laboratory and ensure all recommendations for testing/reporting are followed. The procedures in the manufacturer's product insert takes precedent over those found in CLSI/NCCLS standards.
			Obtain <b>written documentation</b> from medical staff for testing/reporting of organisms/antimicrobial agents beyond those suggested in CLSI/NCCLS standards.
			Add <b>temperature ranges</b> to AST procedures, if appropriate: 33-35°C for staphylococci; 33-37°C for other organism/antimicrobial combinations.
			Review indications for use of <b>daptomycin</b> and develop protocol for testing/reporting with medical staff, if appropriate.
			Review indications for use of <b>telithromycin</b> and develop protocol for testing/reporting with medical staff, if appropriate.
			Review indications for using recommended ATCC strains for <b>quality assessment (QA)</b> . Strains currently suggested for QA include: <i>S. aureus</i> ATCC BAA-976 (D zone test) <i>S. aureus</i> ATCC BAA-977 (D zone test) <i>Klebsiella pneumoniae</i> ATCC 700603 (sometimes recommended for QC also)
<b>Gram Negatives</b>			
			Perform <b>ESBL testing on <i>Proteus mirabilis</i></b> when isolated from significant sources. Note that cefpodoxime, ceftazidime, and cefotaxime are appropriate drugs to screen for ESBL production in <i>P. mirabilis</i> and screening breakpoints for cefpodoxime differ from those for <i>E. coli</i> and <i>Klebsiella</i> spp. Phenotypic confirmatory testing and reporting for <i>P. mirabilis</i> is identical to that for <i>E. coli</i> and <i>Klebsiella</i> spp.
			For <b>ESBL screen test QC</b> , test <i>K. pneumoniae</i> ATCC 700603 OR <i>E. coli</i> ATCC 25922 daily or weekly. Use <i>K. pneumoniae</i> ATCC 700603 for <b>QA</b> .
			For <b>ESBL phenotypic confirmatory test QC</b> , test <i>K. pneumoniae</i> ATCC 700603 AND <i>E. coli</i> ATCC 25922 daily or weekly.
			Review agents tested/reported on <b><i>Pseudomonas aeruginosa</i>, <i>Acinetobacter</i> spp., <i>Burkholderia cepacia</i>, and <i>Stenotrophomonas maltophilia</i></b> and consider modifying procedures based on new recommendations in M100-S15.
			Identify a strategy for testing supplemental agents when an isolate is encountered that is resistant to all drugs on routine test panel. This may include MIC testing of <b>polymyxin B</b> for highly resistant non-Enterobacteriaceae. Reference laboratory assistance may be appropriate.
			Identify a strategy for AST of <b><i>Neisseria meningitidis</i></b> , when requested. Include a safety protocol for handling this species. Reference laboratory assistance may be appropriate.
<b>Gram Positives</b>			
			Review potential utility of the <b>cefoxitin disk diffusion test</b> for <i>mecA</i> -mediated resistance in <b><i>S. aureus</i></b> . Cefoxitin disk diffusion

			test is comparable to OX MIC, OX disk diffusion, PBP2a and <i>mecA</i> for <i>S. aureus</i> .
			Review potential utility of the <b>cefoxitin disk diffusion test</b> for <i>mecA</i> -mediated resistance in <b>coagulase-negative staphylococci (CoNS)</b> . Cefoxitin disk diffusion test is superior to OX MIC and OX disk diffusion for CoNS.
			Determine if <b>testing for <i>mecA</i></b> would be appropriate when equivocal results for oxacillin are encountered in <i>S. aureus</i> or CoNS. This would pertain to isolates from critical sources following discussion with MD. Reference laboratory assistance may be appropriate.
			Apply <i>S. aureus</i> oxacillin MIC and zone diameter breakpoints to <b><i>S. lugdunensis</i></b> . Apply cefoxitin zone diameter breakpoints similarly, if appropriate.
			If using an AST that is unreliable in detecting VISA or VRSA, perform <b>BHI vancomycin screen on <i>S. aureus</i></b> .
			Add CDC's " <b>Algorithm for Testing <i>S. aureus</i> with Vancomycin</b> " to AST procedure for <i>S. aureus</i> .
			Obtain <b><i>S. aureus</i> ATCC BAA-976 and <i>S. aureus</i> ATCC BAA-977</b> for QA of the D zone test for inducible clindamycin resistance.
			Apply new <b>gatifloxacin, levofloxacin and moxifloxacin</b> disk diffusion and MIC breakpoints for <b>staphylococci</b> .
			Discontinue any extrapolation of results among <b>gatifloxacin, levofloxacin and moxifloxacin for <i>Streptococcus pneumoniae</i></b> .
			Perform <b>D zone test</b> for inducible clindamycin resistance on <b>beta-hemolytic streptococci</b> prior to reporting a clindamycin-S result on an erythromycin-R and clindamycin-S isolate. Determine those circumstances when such testing is warranted.
			Implement MMWR 51 (RR-11), 2002 <b>Group B <i>Streptococcus</i> screening</b> procedure for pregnant women to include AST of isolates from women at high risk for penicillin anaphylaxis.
<b>QA/QC</b>			
			Modify <b>QC limits</b> for those organism/antimicrobial agent combinations listed in M100-S15 (boldface type) for which there has been a change in the QC range.
			Test <b>ATCC QC strains</b> daily or weekly even if no new testing reagents/materials have been put into use since last QC testing was performed.
			Follow manufacturer's QC procedures and use the <b>manufacturer's QC ranges</b> even if these differ from CLSI/NCCLS QC limits.
			When submitting <b><i>Salmonella spp.</i></b> to public health authorities, inform them if the isolate demonstrated intermediate or resistant results to a 3 <sup>rd</sup> generation cephalosporin, intermediate or resistant results to a fluoroquinolone and/or resistance to nalidixic acid.
			Review " <b>Summary of Comments and Subcommittee Responses</b> " found on pages 84-86 (M2) and pages 160-162 (M7) in M100-S15.